AMENDMENTS TO THE SPECIFICATION

Please replace the following paragraphs as presented in the specification as most recently filed (on April 21, 2006), with the paragraphs set forth below.

Page 23, paragraph beginning at line 20

FIG. 2C: Nominal 300 ng samples of DNA were aliquoted from a master batch containing surfactant and processed through commercial miniprep columns. Eluate was recycled through Qiaquiek TM-QIAquik® (PCR purification kit) columns and collected either 3 times (4, 5) or twice (6,7) or recycled through Zymoclean (gel DNA recovery kit) columns and collected twice (8,9). Samples were alcohol precipitated using a commercial coprecipitant, electrophoresed on 1.5% agarose gels modified with Synergel (synergistic gelling and sieving agent), stained with SybrGold (nucleic acid gel stain) dye, digitized on a Storm Storm Storm 860 (phosphoimager) and compared to unmodified but reprecipitated samples from the same master batch (10,11). Lanes 1-3: 100,50 and 5 ng of lambda-DNA.

Page 29, paragraph beginning at line 11

Release profiles for hydrophillic hydrophilic dispersed atomized nanocapsules were linear, showed no zero burst and resulted in about 60% release after 72 hours (See Figure 42B). Formula W, manufactured with a standard surfactant (Tween 80) at a reasonable loading value (0.4%) failed to completely release loaded DNA. Figure 2C illustrates that small amounts of DNA (in this case 300 nanograms of DNA) can be recovered accurately in a procedure comprising butanol extraction of 10% butanol/saline releasing fluid followed by isolation on a miniprep column and measurement of absorbance at 260 nm excitation. Results obtained from UV spectroscopy are confirmed by electrophoresis of recovered DNA following alcohol coprecipitation with a commercial coprecipitant aid. Experiment 1A demonstrates the importance of a multi-phase system in creating coated particles from the micellar solution, defines surfactant requirements and validates method for measuring *in vitro* release profiles.